

The Effect of Mating Complexity on Gene Drive Dynamics

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ABSTRACT: Gene drive technology promises to deliver on some of the global challenges humanity faces today in health care, agriculture, and conservation. However, there is a limited understanding of the consequences of releasing self-perpetuating transgenic organisms into wild populations under complex ecological conditions. In this study, we analyze the impact of three such complexities—mate choice, mating systems, and spatial mating network—on the population dynamics for two distinct classes of modification gene drive systems. All three factors had a high impact on the modeling outcome. First, we demonstrate that distortion-based gene drives appear to be more robust against mate choice than viability-based gene drives. Second, we find that gene drive spread is much faster for higher degrees of polygamy. Including a fitness cost, the drive is fastest for intermediate levels of polygamy. Finally, the spread of a gene drive is faster and more effective when the individuals have fewer connections in a spatial mating network. Our results highlight the need to include mating complexities when modeling the properties of gene drives, such as release thresholds, timescales, and population-level consequences. This inclusion will enable a more confident prediction of the dynamics of engineered gene drives and possibly even inform about the origin and evolution of natural gene drives.

Keywords: gene drive, mating complexity, mate choice, mating system, mating network, risk assessment.

Introduction

Gene drive technology is being developed to potentially deliver on some of the critical challenges in human health, agriculture, and biodiversity conservation (Windbichler et al. 2011; Johnson et al. 2016; Prowse et al. 2017; Buchman et al. 2018; Brossard et al. 2019). A prominent example of gene drive is to push transgenes into wild mosquito populations that make them resistant to malarial parasites (Gantz et al. 2015; Carballar-Lejarazú et al. 2020). In conservation studies, the potential of gene drives

to control the spread of invasive species or implement disease resistance in endangered species is being discussed (Johnson et al. 2016; Prowse et al. 2017; Godwin et al. 2019). In agriculture, gene drives could control pest populations, such as fruit flies on cherry plantations, or transform the pest population to make it more susceptible to pesticides (Buchman et al. 2018; Barrett et al. 2019). To date, no gene drive organisms have been released into wild populations. All gene drive constructs are necessarily transgenic and require the release of genetically modified organisms into wild populations. The possibility exists that not all of the unintended consequences of gene drive releases are reversible. Consequently, modeling is key to evaluating this technology.

Theoretical and laboratory studies indicate that some transgenic-driving constructs could spread through wild populations in a relatively small number of generations (Burt 2003; Deredec et al. 2008; Windbichler et al. 2011; Simoni et al. 2020). However, such results may be valid only under ideal conditions, such as random mating and other simplified ecological interactions. Therefore, such estimates may not provide robust predictions of the drive's behavior under field conditions in some circumstances. Several studies related to the risk assessment of gene drives have highlighted the relevance of ecological and technological bottlenecks, such as resistance evolution, mate choice, mating system, and spatial interaction, in successfully deploying gene drive organisms (Oye et al. 2014; National Academies of Sciences, Engineering, and Medicine 2016; Collins 2018; Moro et al. 2018; Giese et al. 2019). Thus, assessing the validity of the model's assumptions is an essential task that any gene drive technology needs to overcome to become an option for a field release. While numerous laboratory assumptions may be violated in the wild, we choose to focus on aspects relating to the ecological complexity of mating. We demonstrate that mate choice, aspects of the mating system, and spatial aspects

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of finding mating partners can change the course of evolutionary trajectories of gene drive systems.

Gene drive leverages sexual reproduction by biasing the inheritance of a specific gene from one generation to the next. Hence, it becomes imperative to account for the target species' reproductive biology and mating pattern to predict critical parameters for a release, such as the threshold of gene drive organisms (GDOs) needed in order to be successful (National Academies of Sciences, Engineering, and Medicine 2016; Moro et al. 2018). While theoretical explorations and laboratory experiments often assume simplified mating conditions based on random mating, other factors also influence mating success in the wild. Nonrandom mating may result from various factors and processes, such as inbreeding, mate choice, and multiple matings, that are often part of complex mating systems. These aspects have already been recognized in gene drive research (Deredec et al. 2008; Unckless et al. 2015; Noble et al. 2017; Qureshi et al. 2019). Inbreeding diminishes the frequency of heterozygotes in the population, slowing the spread of a gene drive (Bull et al. 2019b; Champer et al. 2021). In a natural meiotic drive, females of some species can discriminate against males carrying the drive when the region containing the drive gene is linked to mate choice signals (Price and Wedell 2008; Wedell and Price 2015). For example, the naturally occurring selfish genetic element (*t*-complex) in *Mus domesticus* exhibits mate preference whereby both sexes appear to avoid heterozygous mates by means of olfactory cues (Lenington 1983, 1991; Lindholm et al. 2013).

A newly evolved natural distorter system may remain at low frequency because of reduced fertility of drive-carrying individuals, with the resulting potential to select for mating bias (Charlesworth and Charlesworth 2010; Wedell and Price 2015). However, it remains unclear whether bias in mate preference can quickly evolve for laboratory-engineered gene drives. A study by Drury et al. (2017) showed that nonrandom mating caused by inbreeding could render the CRISPR-based gene drive inefficient against standing genetic variation, resulting in cleavage resistance for Cas9 target sites in the flour beetle *Tribolium castaneum*. Bull (2017) suggested that mild levels of initial inbreeding can lead to the evolution of selfing in hermaphrodites (plants) in response to a homing endonuclease gene drive. Suppression gene drives, aimed at the local eradication of target species, can lead to the evolution of sibmating, significantly hampering the spread of the driven gene (Bull et al. 2019b).

The mating system of target species will also play an essential role in determining the population dynamics of the spread of gene drives. For example, even in the absence of precopulatory mate choice, the *t*-haplotype meiotic drive in mice can be limited by their polyandrous

mating system, where females mate with multiple males in a breeding cycle (Lindholm et al. 2016; Manser et al. 2017). The *t*-haplotype-carrying males have reduced fertility, so when a female mates with multiple males, fertilization by non-drive-carrying males due to sperm competition is more likely (Manser et al. 2017, 2020). A sex-linked gene drive based on utilizing *t*-haplotypes has been proposed to suppress rodent populations (Leitschuh et al. 2018; Godwin et al. 2019). The impact of polyandry on the population-level dynamics of one such proposed gene drive construct (*t*-*Sry*) has been studied by Manser et al. (2019). The construct *t*-*Sry* has two components, *t*-haplotypes and the sex-determining *Sry* gene, and polyandry negatively affects its spread (Manser et al. 2019). Focusing on an age-structured population, Huang et al. (2009) showed that the mating system for *Medea* and engineered underdominance gene drives can significantly change the predicted threshold number of released transgenic individuals for successful population transformation. They also found that low polyandry levels can hamper gene drive spread if only males are released. When the gene drive causes male scarcity (*Y*-shredder), in polygamous systems where males mate with multiple females, the gene spread efficacy is hampered (Prowse et al. 2019).

Most wild populations do not exist in a single panmictic population but often exist as multiple heterogeneous communities. In a spatially segregated population, individuals are more likely to interact with others in their vicinity than randomly with everyone in the population. Some mathematical models of gene drive use numerical evaluation in continuous space to account for such spatial interaction (Beaghton et al. 2017; Tanaka et al. 2017; Girardin et al. 2019). In these systems, the time required for a gene to spread depends on the interaction zone where the wild type meets the transgenics. This zone is the wave's leading edge in reaction-diffusion models (Beaghton et al. 2017; Tanaka et al. 2017; Girardin et al. 2019). In the case of suppression drives, the wave sweeps through the wild population, leaving empty space (Barton and Turelli 2011; North et al. 2013; Bull et al. 2019a). Compared with panmictic models, the suppression drive can be less effective and slow in spatial models (North et al. 2013; Champer et al. 2020, 2021). When considering long-range dispersal, the wild types could occupy the empty space created by the suppression drive, resulting in local cycles of drive eradication and reoccupation by the wild type (Champer et al. 2021). Similar cyclical dynamics are possible for reversal drives released to convert previously established homing drives (Girardin et al. 2019). A question primarily ignored in some of these spatial models concerns the effect of heterogeneous interaction among individuals during mating. For example, the interactions in mathematical models using reaction-diffusion equations are assumed to be homogeneous. The

spread of the gene drives relies on sexual reproduction, which is most likely not spatially or temporally uniform for all individuals in a population. A population structured in a network can help account for the natural heterogeneity in mating success. We use concepts from network theory and build a model to investigate how spatial mating networks could affect the gene drive's spread.

Risk assessors face fundamental challenges when using models in their assessments. First, understanding modeling approaches and the underlying assumptions for complex applications like synthetic gene drives is far from trivial. Second, evaluating the effects of ecological factors on gene drive efficacy is not intuitive. Hence, in general risk, assessment of GDOs will be complex and include more uncertainties than current genetically modified crops designed for release into the environment (Simon et al. 2018). Analogous to other risk assessments (EFSA document on good modeling practice), modeling can be a valuable tool for risk assessment of genetically modified organisms, acknowledging that modeling is complex even for presumably simple questions like the impact of Bt toxins from transgenic maize (EFSA Panel on Plant Protection Products and Their Residues 2014; Fahse et al. 2018; Dolezel et al. 2020). While modeling ecological effects concerning gene drives is still in its infancy (Dhole et al. 2020), much research focuses on efficacy modeling. However, according to risk assessors, the view needs to be much broader than only efficacy.

The population dynamics consequences of mate choice, mating systems, and mating structure for gene drives are crucial in predicting the transgenic constructs' probability and time to fixation as well as the release threshold for invading wild populations. Here, the effects of mate choice and mating systems are studied using deterministic ordinary differential equations. In contrast, the spatial mating structure uses a network model. Although we use different modeling frameworks for different mating complexities, the underlying gene drive model extends from a standard population genetics perspective. Gene drive systems have previously been categorized on the basis of standard terminology—distortion in gamete proportions, fertility selection, and viability selection (Verma et al. 2021). The need for such a common language across the gene drive literature has been extolled by experts and regulators alike (Alphey et al. 2020). Here, we extend this approach by adding a generalizable understanding of the effect of some aspects of mating complexity on gene drive dynamics.

Model and Results

As is typical for a functioning gene drive, we assume a diploid organism whose life cycle consists of three stages: the zygote, adult, and gamete stages. An adult produces

gametes that combine to form a zygote. The zygote grows up to become an adult, and the cycle continues. We also assume that the organisms are diploid with two alleles for the gene of interest, the wild-type allele (W) and the modified allele aimed to be driven (D). Hence, an individual can be one of three genotypes: WW, DD, and WD. Previous work has shown that the gene drive can arise if a drive-carrying genotype undergoes distortion, viability, or fertility selection that acts during the different life stages of an organism (Verma et al. 2021). Hence, one can categorize various gene drive systems on the basis of pre-existing standard population genetics terminology (distortion, fertility selection, and viability selection). Manipulating the strength of these forces via the engineered construct influences the probability of inheritance, giving rise to gene drive (Verma et al. 2021). Gene drives can also be classified into two types on the basis of the purpose of the release: modification and suppression drives. Suppression drives are aimed to reduce or completely eradicate the wild population, while modification drives are intended to replace the wild population with organisms carrying the gene drives. This article focuses on modification drives resulting from distortion and viability selection.

At the gamete level, distortion in gamete ratios can favor the transmission of the drive allele in the heterozygote. Gametes combine to form zygotes, but specific genotypes may become nonviable, distorting the expected Mendelian proportions of the zygotes. It can give rise to meiotic drive (Sandler and Novitski 1957; Lindholm et al. 2016) and CRISPR-based homing endonuclease gene drive (Noble et al. 2017, 2018). The engineered constructs that work principally by manipulating viability selection are those using such zygotic toxin-antidote mechanisms as Medea (Beeman et al. 1992; Ward et al. 2011; Gokhale et al. 2014), inverse Medea (Marshall and Hay 2011), and Semele (Marshall et al. 2011). The general properties of engineered drives classified as such using standard population genetics terminology provide a clean comparative method when evaluating drives for risk assessment and deployment potential (Verma et al. 2021). Here, we subject the target population to three additional factors relevant to field populations—mate choice, mating structure, and mating systems—to understand their effect on gene drive population dynamics (fig. 1).

Mate Choice

We first consider the null case where there is no gene drive and understand how mate choice bias of wild types against transgenics will affect the population dynamics. The mating rate among the wild types is set to 1. Similarly, the mating rate among the drive types is also 1. Mate choice bias in our model is captured by the parameter

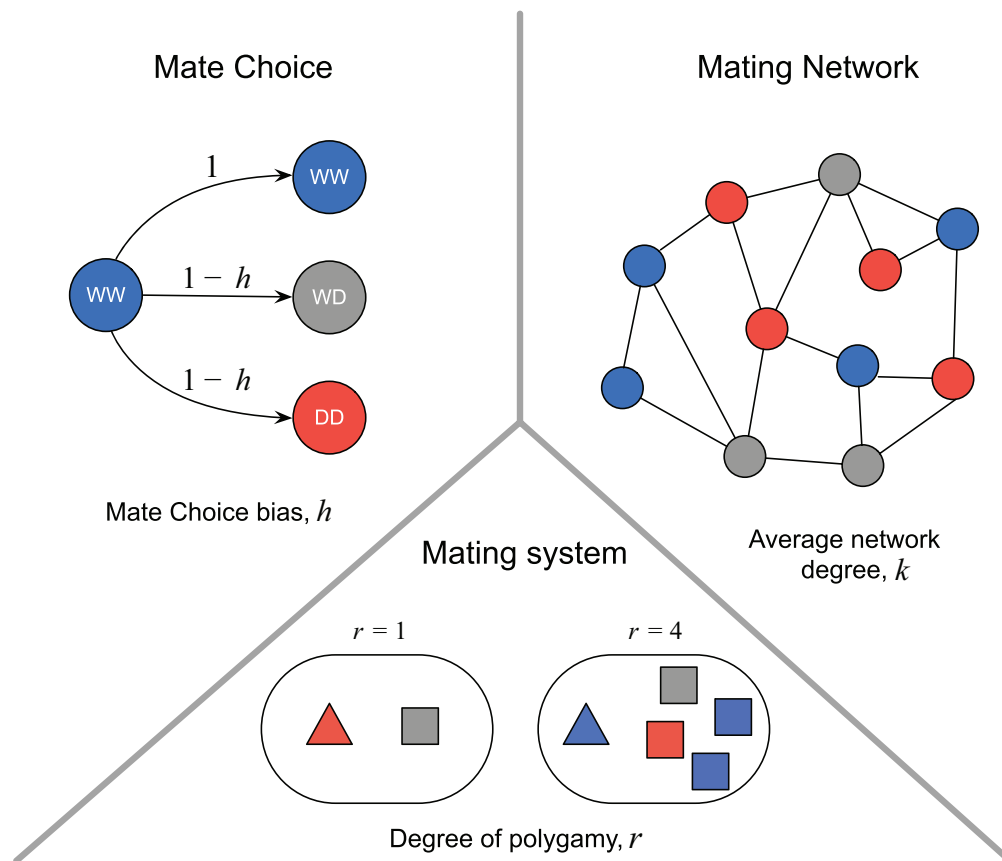


Figure 1: Pictorial representation of the three mating complexities—mate choice, mating network, and mating system—that can affect a gene drive’s population dynamics. Blue, gray, and red colors represent individuals with genotype WW, WD, and DD, respectively. When there is no distinction between the two sexes, individuals are represented by circles, while triangles and squares denote individuals belonging to different sexes. Under mate choice bias, the wild-type genotype (WW) is less likely to mate with a drive-carrying genotype (DD and WD). Mate choice bias is denoted by h in our model, where $(1 - h)$ is the mating rate between the wild types (WW) and the transgenics (WD or DD). In structured mating, individuals mate and reproduce with other receptive individuals in their vicinity, and their likely interactions are modeled in a mating network of average degree k . The consequence of mating with one (monogamy; $r = 1$) or multiple (polygamy; $r > 1$) mating partners for the gene drive dynamics is studied under the mating systems.

h (fig. 1). The mating rate among the wild types (WW) and the drive types (WD or DD) is $(1 - h)$. If $h = 0$, the wild types (WW) are equally likely to mate with the drive-carrying genotypes (WD and DD), while if $h = 1$, the wild types (WW) and the drive types (WD or DD) do not mate at all. During the exploration of parameter space (h), we work under the assumption that the wild-type genotypes are less likely to mate with individuals carrying the drive allele (WD and DD); therefore, $0 \leq h \leq 1$. Our investigation of mate choice is motivated by the theoretical expectation that if variants of ornaments or behaviors that mate choice could act on were linked to the same autosome where the gene drive is located and the cost of the drive is not zero ($c \neq 0$), then mate choice could potentially evolve (Manser 2015). It should be noted that despite substantial speculation and experimentation

with autosomal natural gene drives, particularly the t -haplotype in mice, there is no reliable evidence that mate choice has evolved (Sutter and Lindholm 2016). However, behavioral traits that do appear to be linked to a drive locus have been recently described (Runge and Lindholm 2021). Regardless, it is still potentially valuable to examine the relative sensitivity of various types of gene drive to the impact of mate choice should it evolve. This is not only for perspectives relating to the applied uses of gene drive but also for the origin and evolution of natural drive elements. For simplicity, in our model both sexes (male and female) of WW have an equal bias against mating with WD or DD. The rate of production for the three genotypes, assuming an infinitely large population and random segregation of alleles during meiosis, is given by

$$\begin{aligned}
F_{WW} &= x_{WW}^2 + (1-h)x_{WW}x_{WD} + \frac{x_{WD}^2}{4}, \\
F_{WD} &= (1-h)x_{WW}x_{WD} + x_{WD}x_{DD} \\
&\quad + 2(1-h)x_{WW}x_{DD} + \frac{x_{WD}^2}{2}, \\
F_{DD} &= x_{DD}^2 + x_{WD}x_{DD} + \frac{x_{WD}^2}{4},
\end{aligned} \tag{1}$$

where x_α and F_α are the frequency and rate of genotype production, respectively, and $\alpha \in (WW, WD, DD)$. The following set of differential equations governs the population dynamics of the genotypes in continuous time:

$$\dot{x}_\alpha = F_\alpha - x_\alpha \bar{F}. \tag{2}$$

Here, $\bar{F}$ is the average fitness of the three genotypes:

$$\bar{F} = \sum_{\alpha} F_{\alpha}. \tag{3}$$

The frequencies of all genotypes is normalized to 1:

$$x_{WW} + x_{WD} + x_{DD} = 1. \tag{4}$$

The above constraints on frequencies allow us to represent the dynamics of equation (2) on a de Finetti diagram. The frequency of the three genotypes (WW, WD, and DD) without mate choice ($h = 0$) converge to Hardy-Weinberg equilibrium (Gokhale et al. 2014; Verma et al. 2021). When we introduce the mate choice parameter into the rate equations (1), the dynamics deviate from Hardy-Weinberg equilibrium and are governed by the fixed points that appear in the interior of the de Finetti diagram. In this context, a fixed point is a specific composition of the population ($x_{WW}^*, x_{WD}^*, x_{DD}^*$), where the proportion of all of the genotypes does not change—specifically, where $\dot{x}_\alpha = 0 \forall \alpha \in (WW, WD, DD)$. Primarily, there are two types of fixed points: stable and unstable. If the population is at the stable fixed point, a slight change in the population composition will bring the population to the stable fixed point, while at unstable fixed points, a small change will diverge the population composition away from an unstable fixed point. The position of these fixed points governs the overall population dynamics of a specific case. For example, the population dynamics for a particular case of $h = 0.9$ is shown in the inset of figure 2A. The position of an unstable interior fixed point decides the evolutionary fate of the population.

In figure 2, we plot the positions and trajectories of these interior fixed points for different mate choice (h) values under scenarios such as the null case, viability selection, distortion, and fertility selection. The null case is when only the effect of mate choice is considered without any gene drive arising from viability selection, distortion, and fertility selection (fig. 2A). Even under slight mate choice bias ($h = 0.01$), the dynamics quickly deviate from Hardy-Weinberg equilibrium. An unstable fixed

point (saddle point) appears in the interior of the de Finetti diagram. The threshold frequency of transgenic genotype (DD or WD) required for population transformation is closely related to the position of these unstable fixed points. The area to the left of the unstable fixed point is the basin of attraction of the wild-type genotype. The trajectories of the initial conditions in this area lead to the extinction of the modified allele. In contrast, the area on the right is the basin of attraction of drive homozygotes (DD), leading to population transformation. Increasing the mate choice bias (or as h increases from 0.01 to approximately 1), the position of the interior fixed point moves toward the middle of the WW and DD line (fig. 2A). It implies that when the mate choice bias increases, the threshold amount of transgenics (DD and WD) required to transform the wild-type population increases even without the gene drive.

Mate Choice with Viability Selection (Medea). Many toxin-antidote gene drive designs, including Medea, inverse Medea, Semele, and designed underdominance drive, exhibit viability selection (Beeman et al. 1992; Marshall and Hay 2011; Marshall et al. 2011). In such systems, specific offspring become nonviable during the zygote stage of the life cycle. We have focused on the Medea gene drive system in our analysis, where d measures the drive efficiency. In the Medea gene drive, wild-type homozygous offspring of heterozygous mothers become nonviable (Ward et al. 2011; Akbari et al. 2014; Gokhale et al. 2014; Buchman et al. 2018). The rate of production of genotypes in the Medea gene drive with the incorporation of mate choice bias can be written as

$$\begin{aligned}
F_{WW} &= x_{WW}^2 + (1-h)(1-0.5d)x_{WW}x_{WD} \\
&\quad + (1-d)\frac{x_{WD}^2}{4}, \\
F_{WD} &= (1-h)\frac{x_{WW}x_{WD}}{2} + x_{WD}x_{DD} \\
&\quad + 2(1-h)x_{WW}x_{DD} + \frac{x_{WD}^2}{2}, \\
F_{DD} &= x_{DD}^2 + x_{WD}x_{DD} + \frac{x_{WD}^2}{4}.
\end{aligned} \tag{5}$$

Figure 2B shows the position and trajectory of the unstable fixed point for the viability selection-based Medea gene drive with 100% efficiency (i.e., $d = 1$). The population dynamics equation can be derived using equations (2) and (5). When the mating rate between transgenics and wild types decreases via h , the unstable fixed point moves toward the DD vertex in the de Finetti diagram following a projectile trajectory (fig. 2B). Hence, mate choice bias here increases the threshold release of transgenics. For $h \approx 1$, the number of transgenics released needs to be almost half the target population size for achieving total population

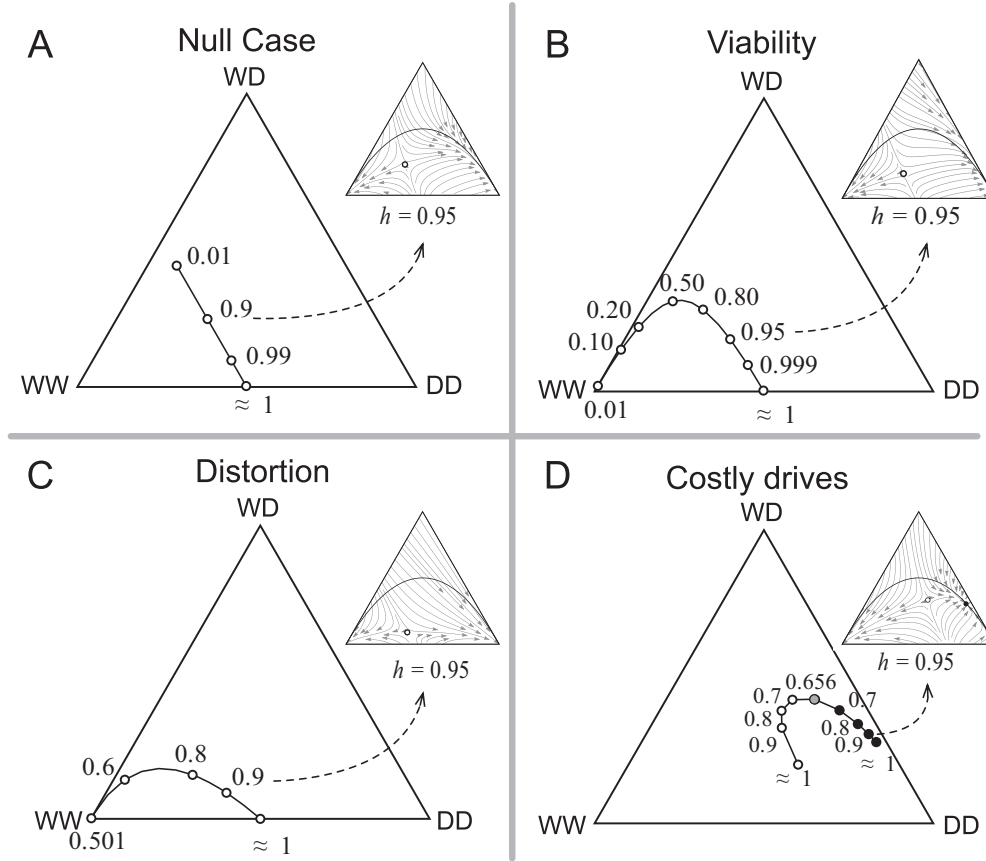


Figure 2: Effect of mate choice bias (h) on the internal fixed point of the population dynamics without (null case) and with gene drive systems based on viability selection (Medea), distortion, and when they are costly. Fixed points appear in the interior of the de Finetti diagrams when the fitnesses of all of the genotypes are the same. Open circles denote unstable fixed points of the dynamics, while closed black circles denote stable fixed points. The gray circle denotes the bifurcation point where both unstable and stable points emerge. The position of these fixed points changes with mate choice bias (h) and hence the overall population dynamics, including the release threshold. Solid black lines show the trajectory of these fixed points for varying mate choice parameter h . A, The null case (without drive) considers the effect of mate choice alone on the population dynamics. B, The Medea drive efficiency is set to 100% ($d = 1.0$). C, The distortion-based drive is assumed to be fully efficient (probability $p = 1.0$). D, The fitness cost of the drive allele is set to $c = 0.2$ and is thus under a multiplicative cost model ($f_{WD} = (1 - c)$ and $f_{DD} = (1 - c)^2$ with $f_{WW} = 1$). When other parameters are not changed, their values are $d = 0$, $p = 0.5$, $f_{WW} = 1$, $f_{WD} = 1$, and $f_{DD} = 1$. The population dynamics for a specific case of h are shown in the insets for all panels.

replacement. These results are also consistent with the invasion condition of equation (A3) derived in appendix A for the Medea gene drive.

Mate Choice with Distortion. In this section we will consider the case of distorted allele transmission in addition to mate choice bias introduced by h . There are several distortion-based gene drives, but here we will focus on a meiotic drive where the distortion efficiency is p . More specifically, p is the probability of transmission of the drive allele from a heterozygous parent to offspring. If $p = 1$, the gene drive system mimics the CRISPR/Cas9-based homing endonuclease drive with 100% efficiency (Noble et al. 2017). If a drive allele is transmitted from

heterozygous parents with probability p , the rate of genotype production then changes to

$$\begin{aligned} F_{WW} &= x_{WW}^2 + 2(1-h)(1-p)x_{WW}x_{WD} + (1-p)^2x_{WD}^2, \\ F_{WD} &= 2(1-h)px_{WW}x_{WD} + 2(1-p)x_{WD}x_{DD} \\ &\quad + 2(1-h)x_{WW}x_{DD} + 2p(1-p)x_{WD}^2, \\ F_{DD} &= x_{DD}^2 + 2px_{WD}x_{DD} + p^2x_{WD}^2. \end{aligned} \quad (6)$$

Again, the population dynamics for the distorted case are given by equation (2), but the effective genotype production rate changes according to equation (6). In figure 2C, we focus on the scenario where the distortion-based gene drive,

such as a meiotic drive or a 100% efficient homing-based CRISPR drive, operates (refer to eq. [6] for $p = 1$). We observe that the interior unstable fixed point appears only after the mate choice bias becomes greater than 50%, or $h > 0.5$, unlike the viability-based gene drive Medea (fig. 2B, 2C). For $h < 0.5$, a small transgenic release is enough for population transformation to drive homozygotes (DD). Hence, the distortion-based gene drives appear to be more robust against mate choice than the viability-based gene drive Medea. These results are also consistent with the condition of invasion derived in appendix A for the distortion-based gene drive (see eq. [A6]).

Costly Drives. The relative number of offspring produced may differ because of the variation in the fertility of the mating pairs resulting from their genotypes, especially if the drive confers a cost. The fitness component due to differential fitnesses is included in the parameters f_α , where $\alpha \in (WW, WD, DD)$. The rate of offspring production for the three genotypes is then given by

$$\begin{aligned} F_{WW} &= f_{WW}^2 x_{WW}^2 + (1-h)f_{WW}f_{WD}x_{WW}x_{WD} + f_{WD}^2 \frac{x_{WD}^2}{4}, \\ F_{WD} &= (1-h)f_{WW}f_{WD}x_{WW}x_{WD} + f_{WD}f_{DD}x_{WD}x_{DD} \\ &\quad + 2(1-h)f_{WW}f_{DD}x_{WW}x_{DD} + f_{WD}^2 \frac{x_{WD}^2}{2}, \\ F_{DD} &= f_{DD}^2 x_{DD}^2 + f_{WD}f_{DD}x_{WD}x_{DD} + f_{WD}^2 \frac{x_{WD}^2}{4}. \end{aligned} \quad (7)$$

To observe the effect of fitness cost, we consider a scenario where $f_{WW} = 1$, $f_{WD} = (1-c)$, and $f_{DD} = (1-c)^2$ for the dynamical equations derived using equation (7). Here, we assume multiplicative fitness cost, where c denotes the fitness cost of the drive allele. The two internal fixed points appear only after substantial mate choice bias ($h \approx 0.656$; fig. 2D). One of the fixed points is unstable, and the other is stable. Therefore, with the multiplicative fitness cost of the transgenic organism, because of drive-allele payload mate choice can result in the coexistence of all three genotypes. When $h < 0.656$, the global stable fixed point lies at the vertex of the wild-type population (WW); hence, no amount of drive release can replace the wild population. However, complete fixation may not be a necessary aim in all applied scenarios.

Besides understanding the impact of mate choice on the population dynamics, we also indirectly probe the threshold release fraction of transgenic organisms required for complete population replacement relative to the target population size. In figure 3, we numerically calculate the threshold frequency of drive homozygotes (DD) necessary to invade a population consisting of wild types (WW). We evaluate the impact of mate choice bias (h), gene drive efficiency, and fitness cost for two gene drive systems, namely,

meiotic drive and Medea. Figure 3A shows that the mate choice bias increases the invasion threshold frequency of DD required for complete population replacement for the Medea drive. The threshold frequency of DD also slightly increases with decreasing drive efficiency. The change in threshold frequency due to drive efficiency reduces for increasing bias in mate choice. The release threshold is close to zero for lower mate choice bias, represented by the heat map's light color. The position of the fixed point for the case of 100% drive efficiency ($p = 1$ and $d = 1$) for both figure 3A and figure 3B corresponds to the scenario studied in figure 2B and figure 2C, respectively. Lower mate choice and sufficiently high distortion probability do not change the threshold frequency for the distortion-based drive. The region in the heat map where a minimal transgenic release can transform the population is significantly higher for the distortion-based drive than for the Medea drive. When the mate choice bias is high enough ($h > 0.5$), an increase in the distortion probability only slightly decreases the invasion threshold of DD. In this regime ($h > 0.5$), a substantial frequency of DD is required for the wild-type population to be invaded even for a very high distortion probability.

Figure 3C and 3D corresponds to the case where there is a fitness cost for the drive-carrying organism ($c = 0.1$; hence, $f_{WD} = 0.9$ and $f_{DD} = 0.81$). Fitness cost leads to an increase in the invasion threshold frequency for both of the gene drive systems overall. Moreover, any DD release is insufficient to invade the wild-type population for inefficient drives under low mate choice bias. The dark color represents this region in the heat map. Interestingly, increasing the mate choice bias can facilitate the invasion by DD even for less efficient drives. The distortion-based gene drive appears to be more robust against the ecological stress of mate choice bias even when considering the fitness costs.

Mating Systems

Gene drive technology relies on sexual reproduction between the mating pairs. Most of the target species of interest have a polygamous mating system instead of the commonly assumed monogamous mating system (Moro et al. 2018; Rode et al. 2019). As introduced in the previous section on mate choice, the model is modified here to incorporate this aspect of the mating system. In this model, we will consider two separate populations of the two sexes. We assume that the offspring of both sexes are produced in equal proportion. The frequency of male and female genotypes are denoted using x_i and y_j . There are three possible genotypes: wild type (WW), drive heterozygotes (WD), and drive homozygotes (DD). Let us consider the mating system when one male mates with r females.

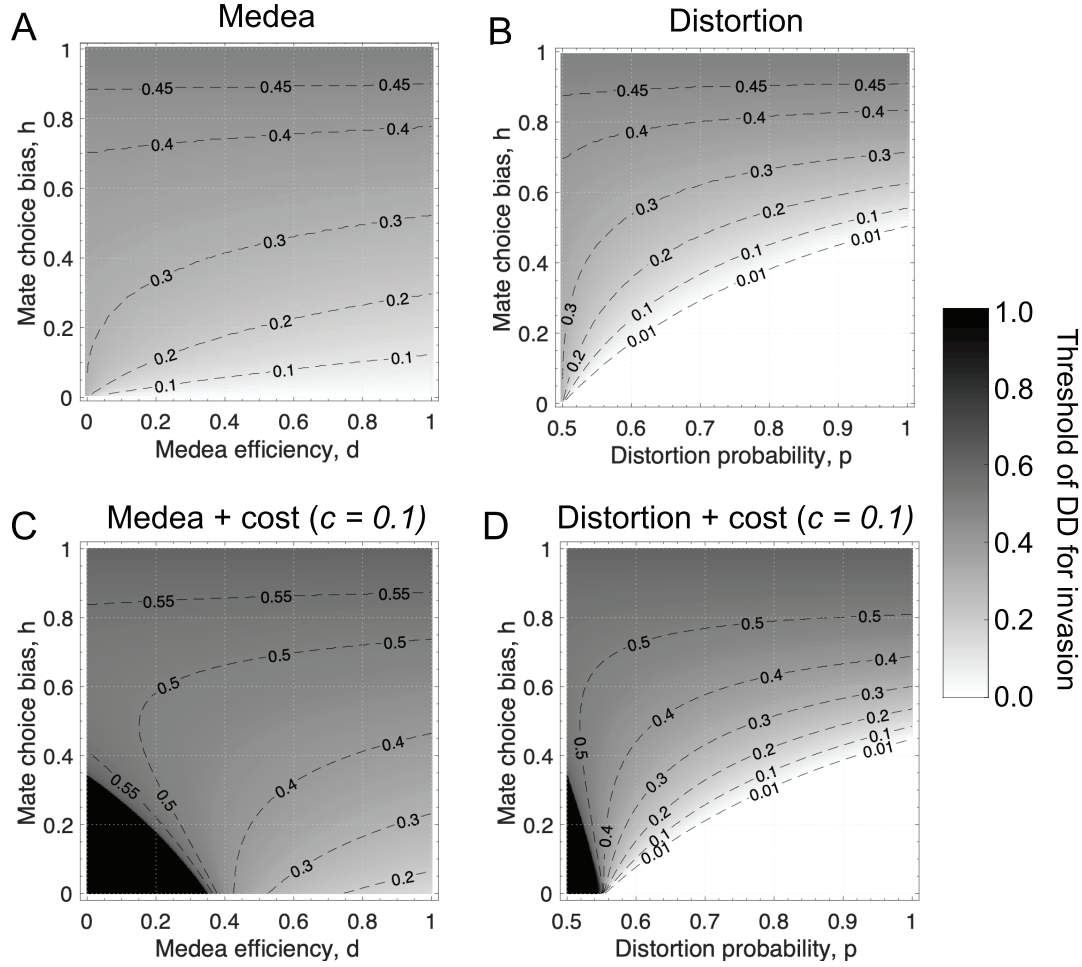


Figure 3: Heat map showing the threshold frequency of drive homozygotes (DD) required to invade a population of wild-type homozygotes (WW) with respect to variation in mate choice bias (h) for the following gene drive systems: Medea and distortion-based drive. Black dashed lines correspond to the contour lines showing the threshold frequency of drive homozygotes (DD). A, Medea gene drive with no fitness cost (i.e., $c = 0$). B, Distortion-based gene drive with no fitness cost to drive (i.e., $c = 0$). C, Medea gene drive where the fitness cost due to the drive allele is $c = 0.1$ (hence, $f_{WD} = 0.9$ and $f_{DD} = 0.81$). D, Distortion-based gene drive where the fitness cost due to the drive allele is $c = 0.1$ (hence, $f_{WD} = 0.9$ and $f_{DD} = 0.81$).

Hence, $r = 1$ represents the monogamous mating system, while $r > 1$ corresponds to the polygynous mating system. The following set of equations gives the frequencies of the genotypes produced with the polygamous mating system, as the equation holds for both males and females (with a change in variable x_i and y_j):

$$F_k(r) = \sum_{\alpha, \beta_1, \beta_2, \dots, \beta_r} \sum_{j=1}^r M_k(\alpha, \beta_j) f_{\alpha} x_{\alpha} \prod_{i=1}^r f_{\beta_i} y_{\beta_i}. \quad (8)$$

Here, $M_k(\alpha, \beta_j)$ is the proportion of genotype k produced from the mating between a male of genotype α and a female of genotype β_j . The terms α and β_j are dummy indexes for any of the three genotypes WW, WD, or DD. The elements

of the matrix $M_k(\alpha, \beta_j)$ will depend on the gene drive type as well. Matrix M_k for Medea (eqq. [A7]–[A9]) and the distortion-based gene drive system (eqq. [A10]–[A12]) is given in appendix A. The summation over α and β_j is carried out over the set of all genotypes (WW, WD, DD). We have also assumed a polygamous mating system of mating ratio r (i.e., one male mates with r female or vice versa). Equation (8) may be interpreted in part as selecting a male of genotype α and selecting r females of genotype $\beta_1, \beta_2, \dots, \beta_r$. Finally, the contribution of all possible matings in producing genotype k is summed up.

Simplifying equation (8) by means of the expansion formula for multinomial expression yields

$$F_k(r) = r F_k(1) (f_{WW} y_{WW} + f_{WD} y_{WD} + f_{DD} y_{DD})^{r-1}. \quad (9)$$

The following set of differential equations governs the population dynamics of the genotypes in continuous time:

$$\begin{aligned}\dot{x}_\alpha &= \frac{1}{2}F_\alpha(r) - x_\alpha\bar{F}(r), \\ \dot{y}_\alpha &= \frac{1}{2}F_\alpha(r) - y_\alpha\bar{F}(r).\end{aligned}\quad (10)$$

Here, $\bar{F}$ is the sum of rates of genotype production:

$$\bar{F}(r) = \sum_{\alpha} F_{\alpha}(r). \quad (11)$$

The total population of both males and females remains constant and sum up to unity:

$$x_{WW} + x_{WD} + x_{DD} = 1, \quad (12)$$

$$y_{WW} + y_{WD} + y_{DD} = 1. \quad (13)$$

In equation (9), the production rate of genotype k is $F_k(r = 1)$ and $F_k(r > 1)$ for monogamous ($r = 1$) and polygamous ($r > 1$) mating systems, respectively. It implies that the equilibrium population dynamics for both monogamous ($r = 1$) and polygamous ($r > 1$) mating systems, even with gene drives, are equivalent. In other words, the final population composition of the genotypes remains the same for both polygamous and monogamous mating systems. Previous studies without any gene drive also support that the equilibrium dynamics for monogamy and polygamy remain the same (Karlin 1978; O'Donald 1980). However, the difference lies in the relative time to reach population equilibrium. It can be shown that after simplifying the equation (10) obtained for $r > 1$, the rate of increase of different genotypes is equivalent to the case of monogamy ($r = 1$) with rescaled time. The expectation is that the gene drive will spread faster in polygamous mating species compared with monogamy (Moro et al. 2018). Hence, the time required for the drive allele to spread through the population should increase for the monogamous mating system. Our result also supports the expected outcome. Here, we quantify the same.

We first look at the case where there is no fitness cost of the gene drive and only the efficiency of the two gene drive systems based on distortion and viability selection are varied. In figure 4, the left and center panels show that the gene drive will spread faster for species with a high degree of polygamy (r). In the material provided on Zenodo (<https://doi.org/10.5281/zenodo.6624757>; Verma 2022) for figure 4, we show that the distortion-based gene drive will spread faster than the viability-based drive. The time for the gene drive to reach 99% frequency is an order of magnitude higher for the Medea drive than for the CRISPR homing drive or the meiotic drive. A higher degree of polygamy (r) reduces the time required to reach critical drive frequency (99%) for both gene drive systems.

The figure 4 panels for costless drives show that the relative time required for the drive allele to reach 99% frequency is rescaled exactly by a factor of $1/r$ for the polygamy relative to the monogamous mating system (e.g., time relative to monogamy is 0.2 for degree of polygamy $r = 5$). This is in line with the relation obtained in equation (9). When $f_{WW} = f_{WD} = f_{DD}$, the production rate of offspring for polygamy is r times that for the monogamous mating system. But when we have a fitness cost c for carrying a drive allele, the relation between the time to reach 99% frequency and the degree of polygamy becomes more complex (fig. 4). An increase in the degree of polygamy first decreases the relative time to reach the drive allele's critical frequency ($r = 2$ and $r = 4$), but a further increase in the degree of polygamy ($r = 6, 8, 10$) elevates it. In figure 4, it can also be noted that when the distortion probability is low ($p < 0.625$), the costly drive allele is not able to invade the wild-type population. Thus, in the right panel we explore values of distortion probability above this threshold. This is in congruence with the condition of invasion derived for the monogamous case in equation (A6).

The above result can be understood from equation (9), where the fitness cost makes the factor $(f_{WW}y_{WW} + f_{WD}y_{WD} + f_{DD}y_{DD})^{r-1}$ less than 1. The factor $(f_{WW}y_{WW} + f_{WD}y_{WD} + f_{DD}y_{DD})^{r-1}$ decreases exponentially with increasing level of polygamy r . Hence, the time is effectively rescaled by a factor of $1/(r(f_{WW}y_{WW} + f_{WD}y_{WD} + f_{DD}y_{DD})^{r-1})$. The time first decreases when dominated by $1/r$ with an increase in r but later on decreases when dominated by $1/(f_{WW}y_{WW} + f_{WD}y_{WD} + f_{DD}y_{DD})^{r-1}$. When the fitness cost is $c = 0.2$, the relative time until the drive allele reaches 99% frequency with respect to monogamy decreases for $r = 2$ and $r = 4$, but then it starts to increase for $r = 6$. For $r = 8$ and $r = 10$, the spread of the gene drive becomes slower compared with that of monogamy. Another way to understand the results is that the rate of production genotype DD first increases up to a point for increasing levels of polygamy r but later decreases for moderate fitness cost (fig. B1). Hence, the time needed to spread the gene drive is lowest for intermediate levels of polygamy. Further increase in the degree of polygamy reduces the production of DD and therefore increases the time needed to spread the drive allele.

Spatial Network Interaction

The population dynamics of the CRISPR-based homing endonuclease gene drive have been extensively studied for well-mixed infinitely large (Noble et al. 2017) and finite (Noble et al. 2018) populations. But most species occur in a partially heterogeneous landscape where they interact and mate with other individuals in their vicinity. Hence,

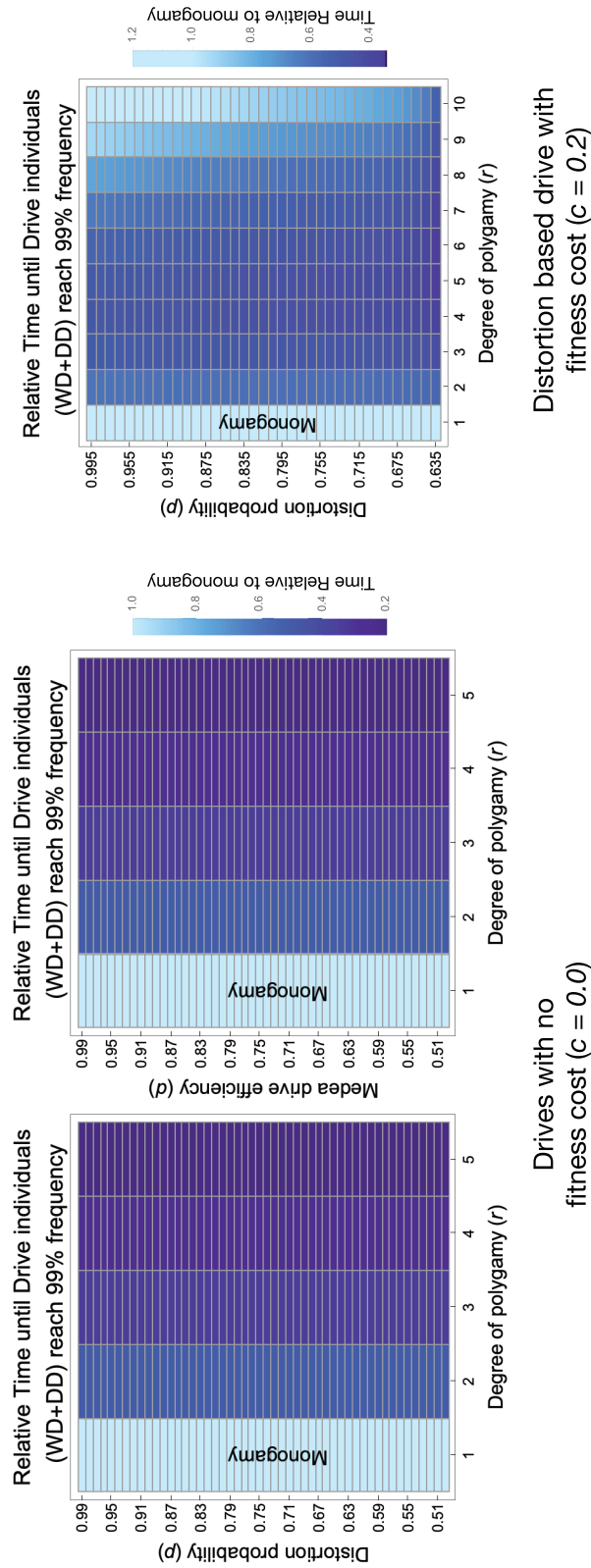


Figure 4: Effect of mating system and drive efficiency on the time for the drive allele to reach 99% frequency. We start from a population consisting of 99% wild types (WW) and 1% drive heterozygotes (DD). The population is evolved until the frequency of drive allele-carrying individuals (WD + DD) reaches 99% (“fixation”). The left and center panels do not include a fitness cost attributable to the drive allele. The left panel shows the fixation time for different degrees of polygamy across the range of Medea efficiency normalized by the time taken in the monogamy case. The center panel shows the fixation time for different degrees of polygamy across the range of distortion efficiency normalized by the time taken in the monogamy case. Both panels show a linear dependence on the degree of polygamy, which continues for higher values of r (not shown; available on Zenodo [https://doi.org/10.5281/zenodo.6624757; Verma 2022]). When including a fitness cost for carrying a drive allele ($c = 0.2$), the fixation time shows a nonlinear dependence on the degree of polygamy. For low and high degrees of polygamy, it takes longer to reach fixation than for intermediate levels of polygamy. There is also a slight dependence on the efficiency of the distortion drive, where a lower probability of distortion is still faster for intermediate levels of polygamy compared with the monogamous case.

a network-based population is an appropriate framework to model dynamics in such structured populations.

We considered a structured population of n individuals. The individuals live on a random network with an average degree of k ; thus, each individual has k connections on average. Here, k controls the number of mating opportunities and the level of competition for an individual. The population is updated via a death-birth process (fig. 5), described as follows. First, an individual is chosen randomly for death. Then two parents are selected, which are neighbors of the dead individual with a probability proportional to their fertility fitness. According to their genetic archetype, the selected parents contribute their gametes, where other genetic effects, such as distortion, can come into play. The combination of these contributed gametes forms the offspring that replaces the dead individual in the network. The population is updated until it fixes to all WW or all DD states.

In figure 6, we exhibit the stochastic network model by running the simulation several times and plotting fixation probability and conditional fixation time with variation in the average number of interacting individuals per site (represented by k). We also studied the impact of increasing the number of released transgenics (WD and DD) and different genotypes (WD and DD). Here, k controls the number of mating opportunities and competition during the birth process. When k increases, the fixation probab-

ity of DD decreases, mainly due to higher competition during the birth update per site (fig. 6A). As expected, distortion probability has a positive impact on the fixation probability of DD. The effect is more pronounced for lower values of an average degree, since the heterogeneity in the number of connected individuals is also high for this case. Fixation probability also increases as the number of released DD transgenics increases (fig. 6A). Releasing DD transgenics has a lower chance of getting fixed than a WD release (fig. 6B). This is because the fitness cost of DD is relatively high compared with that of WD ($f_{WD} = 0.50$, $f_{DD} = 0.25$). If the fitness cost is negligible and the drive efficiency is high, the release of the DD genotype is expected to fix the gene drive with a higher probability. The effect on fixation probability by the release of WD compared with DD becomes more pronounced with the increase in average degree k (fig. 6B). It increases first with an increase in the released number of transgenics, attains a maximum, and decreases later. We present the fixation probability for a gene drive with a high fitness cost in part motivated by observations that many, but not all, natural autosomal gene drives (e.g., *Mus musculus* *t*-haplotype and the *Drosophila melanogaster* segregation distorter [SD] alleles) are frequently homozygous male sterile. However, the impact of the average degree of the network is indeed a result of the cost, as it brings about a density-dependent aspect to the number of released individuals. The effect is clarified

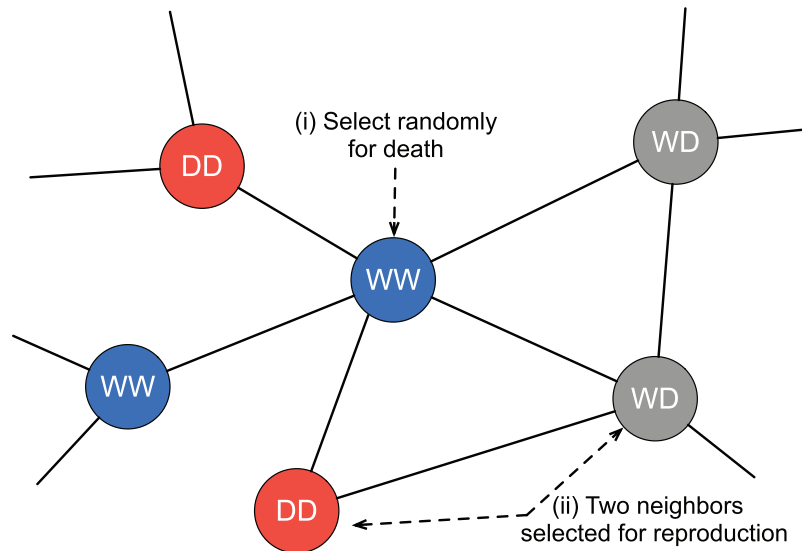


Figure 5: Spatial model explaining the population update mechanism. Blue, gray, and red colors represent individuals of WW, WD, and DD genotypes, respectively. The population update happens in two steps. First, a random individual is selected for death. This step creates space at that particular network position. Second, two random neighbors of the dead individual are chosen as parents to produce offspring. The genotype of the offspring is determined from the parents, and it replaces the dead individual.

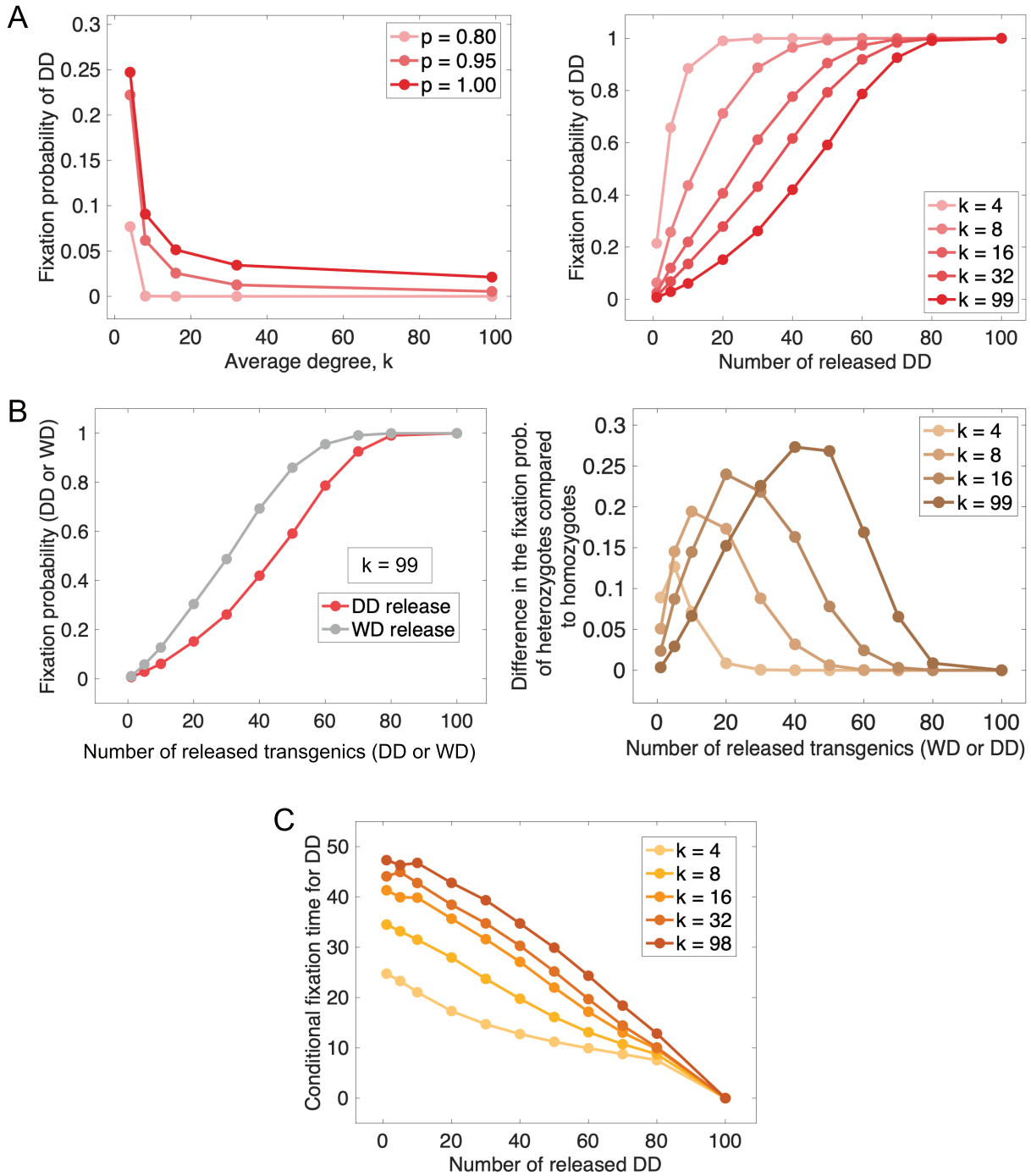


Figure 6: Fixation probability and conditional fixation time of DD with variation in average degree k , distortion probability p , and initial number of released transgenic individuals WD or DD. *A*, Plots showing the fixation probability of drive homozygotes starting from a single DD individual against average degree k (*left*) and number of released DD (*right*) for different values of p and k , respectively. *B*, Fixation probability plotted against the number of released DD and WD for a complete graph ($k = 99$; *left*) and the difference between the fixation probability of WD and DD release plotted against the number of released transgenics for varying average degree k (*right*). *C*, Average number of generations when the drive individuals get fixed in the population against an initial number of released DD with varying average degree k . A generation consists of n death-birth steps. Hence, in a generation the whole population is updated on an average. All simulations use a population size of $n = 100$ and 10,000 trials to estimate fixation probability and conditional fixation time. The distortion probability and fitness cost are fixed to $p = 0.95$ (except *A*, *left*) and $c = 0.5$, respectively. In all our simulations, the release nodes of transgenics are chosen at random.

in figure B2, where we show the importance of discerning the fitness costs of a drive before releasing it into a sparsely or densely connected network.

We also plotted conditional fixation times against the size of transgenic (DD) releases for various values of network degree (fig. 6C). Unsurprisingly, this confirmed that the conditional fixation time is fastest for larger transgenic releases. Interestingly, conditional fixation time is slowest in populations with substantial structure (lower values of k). The broad range of values of conditional fixation times for the smaller release sizes reflects the increasing probability of stochastic loss of D alleles as population structuring increases.

Discussion

Gene drive is one of the tools of synthetic biology that has the potential to transform whole wild populations. The transformation exploits and modulates one of the foundational tenets of evolution—the inheritance of traits through sexual reproduction. Thus, variation in the reproductive biology and mating behavior of the target species can affect the eventual spread of the gene drive. The analytical approach and terminology used in this study were selected to facilitate comparison between different drive mechanisms within a common comparable framework (Verma et al. 2021). While previous studies emphasized the evolution of resistance to gene drives (Price et al. 2020), we have examined some of the ecological assumptions related to mating systems and their effect on the potential outcomes of a drive release (Noble et al. 2017; Unckless et al. 2017; Champer et al. 2018). These include factors related to life histories and social interactions relevant under field conditions, namely, mate choice, multiple matings, and spatial aspects of the mating network. Our work focuses on single-locus gene drives located on autosomes in diploid organisms. From an applied perspective aimed at using gene drives to manipulate populations, this equates to modification drives (not suppression drives, which seek to reduce population sizes). We describe circumstances where the explored factors can substantially influence predictions for the release thresholds of transgenics required for a successful invasion and the possible fixation times.

First, we considered a monogamous situation that deviates from panmixis as individuals actively choose partners on the basis of the presence or absence of a transgenic drive allele D (mate choice). For any drive linked to ornaments or behaviors that could form the basis of heritable mate choice, a mate choice bias can be envisioned with a significant effect on the release threshold of a gene drive, as shown in figure 3. Inefficient drive and fitness

costs due to drive payload aggravate the situation. The predicted threshold release is drastically different from a situation with no mate choice bias (fig. 2). This finding is important to estimate the drive efficacy and is highly relevant for the risk assessment of the drive. Comparing different drive approaches, we found that distortion-based gene drives fare much better than drives based on viability selection under the potentially more ecologically realistic conditions of mate choice. While the regulatory importance of these insights may be limited because of the often unknowable capacity for mate choice to evolve, they may inform what parameters might be helpful to monitor subsequent to any release.

Next, we considered the potential for multiple matings for both females and males. Under the explored scenarios, the final evolutionary outcome of the spread of the gene drive (distortion or Medea drive) for polygamous mating was the same in our simulations as that for the monogamous system. Even the species with a higher degree of polygamy will converge to the same evolutionary fate for a given gene drive system. However, the time needed for the spread of the drive gene will be affected by the level of multiple matings and the fitness cost linked to the drive. Time to fixation will be smaller for a higher degree of polygamy without any fitness cost (fig. 4). However, a moderate fitness cost under different polygamy levels will trigger a nonlinear outcome for the time until drive establishment. This nonlinearity occurs because the production rate of a drive homozygote first increases but later decreases in line with the degree of polygamy for moderate fitness cost (fig. B1). Hence, the drive gene is expected to spread faster for species with intermediate levels of polygamy when there is an associated fitness cost of the drive allele. Regarding mice, which are one of the focus organisms of gene drive applications, these primarily utilize suppression drives that are not considered here. It is, however, still interesting to note that the value of $r = 2$ that is applied for island mice (Birand et al. 2022) would imply a nearly twofold difference in fixation time compared with situations of monogamy, which may be significant in circumstances where alternative nongenetic approaches may be available to achieve the same goals.

Last, we examined the spatial implications of finding mating partners (mating opportunities) on the model outcome. To this end, the framework developed for the spatial mating interaction can be applied to any diploid population, regardless of the presence of a gene drive. Considering a finite population on a network allows us to understand the probable outcomes of gene drive release. A finite population leads to stochastic fluctuations in the frequencies of the genotypes resulting in different outcomes for the same initial conditions. We found that the spread of transgenic release is lowered when individuals, on average, have more

mating opportunities and intrasexual competition. Thus, the fixation time for the transgenic increases with an increase in the average degree of the mating network. Concerning the question of how the connectivity of mating networks varies in wild populations, it is reported that selective pressures under which species evolve shape their network structure in the environment (Pinter-Wollmann et al. 2014). Hence, changes in environmental conditions such as resource availability, seasonal effects, selective pressure, and life history traits can impact the network structure. Within a species, variation at the individual level can also lead to heterogeneous connectivity. Species with sparsely connected individuals on the mating network have a higher chance of fixing drive genes in a shorter time. We also observe that the success in fixation of drive homozygotes can be mitigated by releasing more transgenic individuals. Furthermore, when the fitness cost associated with carrying a drive allele payload is high, releasing drive heterozygotes instead of homozygotes would result in a higher chance of gene drive fixation (fig. 6B). Our model of network dynamics uses a death-birth process. Alternatively, it is also possible that we first select a connected mating pair for birth that replaces an individual connected to either parent for death. In models of asexual reproduction in networks, such birth-death versus death-birth models can show qualitative differences in fixation properties (Zukewich et al. 2013; Hindersin and Traulsen 2015; Kaveh et al. 2015). Exploring such alternative processes would require a thorough understanding of the network dynamics of the target species.

While our findings can collectively be viewed solely in applied uses of synthetically engineered drives, they may also inform the origin and evolution of natural gene drives in animals. Despite the intense focus on synthetic gene drive and analogies with described gene drive elements from nature, three natural elements fit the modeling described here: autosomal in a diploid organism, single copy, and do not alter population sex ratios or involve genomic transposition. The three natural drive elements are described in table 1. In contemporary wild populations, these drives can be detected as polymorphic (with a fre-

quency less than 1). None are associated with mate choice. With the strong caveat that three is a minimal sample size, there is an absence of information on how many drive elements may have evolved and risen to fixation in species (rendering them effectively undetectable except by inter-specific crossing). It is still interesting to speculate whether the findings of this study provide any insight into the origin of natural gene drives and the probability that they spread once they arose in the first individual. Our study would indicate that the greatest probability of a new gene drive allele increasing from a low initial frequency would be in species with low degrees of polygamy and a highly structured population (low k). This would particularly be the case for the *Drosophila melanogaster* SD alleles and mouse *t*-haplotypes, where the cost of the drive (c) appears to be high. To gain confidence in these statements, it will be necessary to estimate the relative frequency of gene drive elements in a range of animals with different ecological properties. However, given the historical difficulty of detecting autosomal natural gene drive elements, it may take some time, even with the drop in genotyping costs, for extensive drive studies to take place outside intensively studied model organisms.

In this study, we decided to focus on three factors related to the mating complexities of the target species. Still, many other ecological and environmental factors can impact the spread of gene drives. Known factors include the age structure of a population, spatial landscape, and seasonality (Huang et al. 2009, 2011; North et al. 2013, 2019, 2020; Eckhoff et al. 2017). Gene drive behavior and the interactions of a drive released in a complex ecosystem over long time periods are highly complex. Navigating this ecological complexity may seem insurmountable (Levin 2003). However, we will face a similar control problem for any technology aiming to intervene in complex systems. As such, it is not workable to address in silico all possible ecological and evolutionary pressures and scenarios that an engineered system will meet in the real world (Denton and Gokhale 2019; Lindvall and Molin 2020). Undoubtedly, modeling will play a key role in understanding drive spread. Our study emphasizes that modeling needs a

Table 1: Examples of natural drive elements with properties simulated in this study

Species	Drive element	Drive type and parameters	Ecological properties
<i>Tribolium castaneum</i>	Medea	Viability drive d	Probably low k , highly polyandrous (Pai and Yan 2020)
<i>Drosophila melanogaster</i>	Selfish segregation distorter (SD)	Distortion drive p, c	Probably low k , polyandry with some remating latency (Singh and Singh 2004)
<i>Mus musculus</i>	<i>t</i> -haplotype	Distortion drive p, c	Probably low k , polyandry ($r = 2$; Birand et al. 2022)

more ecological reality to predict drive behavior. Identifying and collecting necessary information on the effect of primary ecological and evolutionary pressures will thus be crucial to access the risk before any field deployment (National Academies of Sciences, Engineering, and Medicine 2016; James et al. 2018; Long et al. 2020).

Conclusion

Most gene drive modeling focuses on drive spread under simplified conditions, such as panmixis. In this study, we tested whether more complex assumptions, characteristic of many species and mating systems, justify the use of simplified assumptions. The results show that ecological factors related to mating can substantially change drive spread and, as supposed for many other ecological factors, may strongly impact the temporal and spatial dynamics of gene drive systems. Modeling may be used to predict gene drive spread and thus to assist in risk assessment. In this case, mating-related parameters, as with all critical assumptions related to the ecology of the species, need to undergo a reality check. In a wider sense, the new modeling framework, including tools for analyzing spatial interactions or multiple matings, is generic and has the potential to be applied to any diploid population, independent of gene drive applications.

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Statement of Authorship

P.V. and C.S.G. developed the model and performed the model analysis and coding simulations. All authors conceived the project, developed the theory, and wrote the article. All authors read and approved the final manuscript.

Data and Code Availability

All data and simulation code for generating figures are available on Zenodo (<https://doi.org/10.5281/zenodo.6624757>; Verma 2022).

APPENDIX A

Additional Methods

Invasion Condition for the Medea Drive with Mate Choice (h)

If we consider the case of the Medea gene drive with fertility selection, the rate of production of the three genotype is given by the combination of equations (5) and (7):

$$\begin{aligned} F_{WW} &= f_{WW}^2 x_{WW}^2 + (1-h)(1-0.5d)f_{WW}f_{WD}x_{WW}x_{WD} \\ &\quad + (1-d)f_{WD}^2 \frac{x_{WD}^2}{4}, \\ F_{WD} &= (1-h)f_{WW}f_{WD} \frac{x_{WW}x_{WD}}{2} + f_{WD}f_{DD}x_{WD}x_{DD} \\ &\quad + 2(1-h)f_{WW}f_{DD}x_{WW}x_{DD} + f_{WD}^2 \frac{x_{WD}^2}{2}, \\ F_{DD} &= f_{DD}^2 x_{DD}^2 + f_{WD}f_{DD}x_{WD}x_{DD} + f_{WD}^2 \frac{x_{WD}^2}{4}. \end{aligned} \quad (A1)$$

The rate of change of frequencies of each genotype is still given by equation (2). We use the constraint on the frequencies of the three genotypes in equation (4) to reduce the population dynamics of the genotypes to two variables after replacing $x_{WD} = 1 - x_{WW} - x_{DD}$ in equation (2). The drive will not invade the wild-type population if both of the eigenvalues of the dynamical system are negative. Eigenvalues can be deduced from the Jacobian matrix (J_d) of the system at $(x_{WW}, x_{WD}, x_{DD}) = (1, 0, 0)$:

$$J_d = \begin{pmatrix} f_{WD}f_{WW}(1-h) - f_{WW}^2 & f_{WD}f_{WW}(1-h) - 2f_{DD}f_{WW}(1-h) \\ 0 & -f_{WW}^2 \end{pmatrix}. \quad (A2)$$

Hence, the Medea gene drive can invade a population of wild types if

$$(1-h)f_{WD} > f_{WW}. \quad (A3)$$

Note that the above invasion condition is independent of the efficiency of the Medea gene drive (d). This condition is plotted in figure A1A.

Invasion Condition for the Distortion Drive with Mate Choice (h)

Consider the scenario of the distortion-based gene drive with fertility selection. The rate of production of the

three genotypes will then be governed by the combination of equations (6) and (7):

$$\begin{aligned}
 F_{WW} &= f_{WW}^2 x_{WW}^2 + 2(1-h)(1-p)f_{WW}f_{WD}x_{WW}x_{WD} \\
 &\quad + (1-p)^2 f_{WD}^2 x_{WD}^2, \\
 F_{WD} &= 2(1-h)pf_{WW}f_{WD}x_{WW}x_{WD} \\
 &\quad + 2(1-p)f_{WD}f_{DD}x_{WD}x_{DD} \\
 &\quad + 2(1-h)f_{WW}f_{DD}x_{WW}x_{DD} \\
 &\quad + 2p(1-p)f_{WD}^2 x_{WD}^2, \\
 F_{DD} &= f_{DD}^2 x_{DD}^2 + 2pf_{WD}f_{DD}x_{WD}x_{DD} + p^2 f_{WD}^2 x_{WD}^2.
 \end{aligned} \tag{A4}$$

Similar to the Medea gene drive scenario, the population dynamics of the above system can be written in the form of two variables, x_{WW} and x_{DD} , using equation (4). The Jacobian matrix (J_m) of the system at $(x_{WW}, x_{WD}, x_{DD}) = (1, 0, 0)$ is given by

$$J_d = \begin{pmatrix} 2f_{WD}f_{WW}(1-h)p - f_{WW}^2 & 2f_{WD}f_{WW}(1-h)p - 2f_{DD}f_{WW}(1-h) \\ 0 & -f_{WW}^2 \end{pmatrix}. \tag{A5}$$

From the condition on the eigenvalues, the gene drive can invade the wild-type population if

$$2(1-h)pf_{WD} > f_{WW}, \tag{A6}$$

as shown in figure A1B and A1C. Note that when there is no mate choice ($h = 0$), the above condition reduces to the invasion condition derived by Noble et al. (2017) for the CRISPR gene drive.

$M_k(\alpha, \beta_i)$ in equation (8) for the Medea and Distortion-Based Gene Drives

Medea Gene Drive

$$M_{WW} = \begin{bmatrix} 1 & 0.5(1-d_m) & 0 \\ 0.5 & 0.25(1-d_m) & 0 \\ 0 & 0 & 0 \end{bmatrix} \tag{A7}$$

$$M_{WD} = \begin{bmatrix} 1 & 0.5(1-d_m) & 0 \\ 0.5 & 0.25(1-d_m) & 0 \\ 0 & 0 & 0 \end{bmatrix} \tag{A8}$$

$$M_{DD} = \begin{bmatrix} 0 & 0 & 0 \\ 0 & 0.25 & 0.5 \\ 0 & 0.5 & 1 \end{bmatrix} \tag{A9}$$

Distortion-Based Gene Drive

$$M_{WW} = \begin{bmatrix} 1 & (1-p) & 0 \\ (1-p) & (1-p)^2 & 0 \\ 0 & 0 & 0 \end{bmatrix} \tag{A10}$$

$$M_{WD} = \begin{bmatrix} 0 & p & 1 \\ p & 2p(1-p) & (1-p) \\ 1 & (1-p) & 0 \end{bmatrix} \tag{A11}$$

$$M_{DD} = \begin{bmatrix} 0 & 0 & 0 \\ 0 & p^2 & p \\ 0 & 0 & 1 \end{bmatrix} \tag{A12}$$

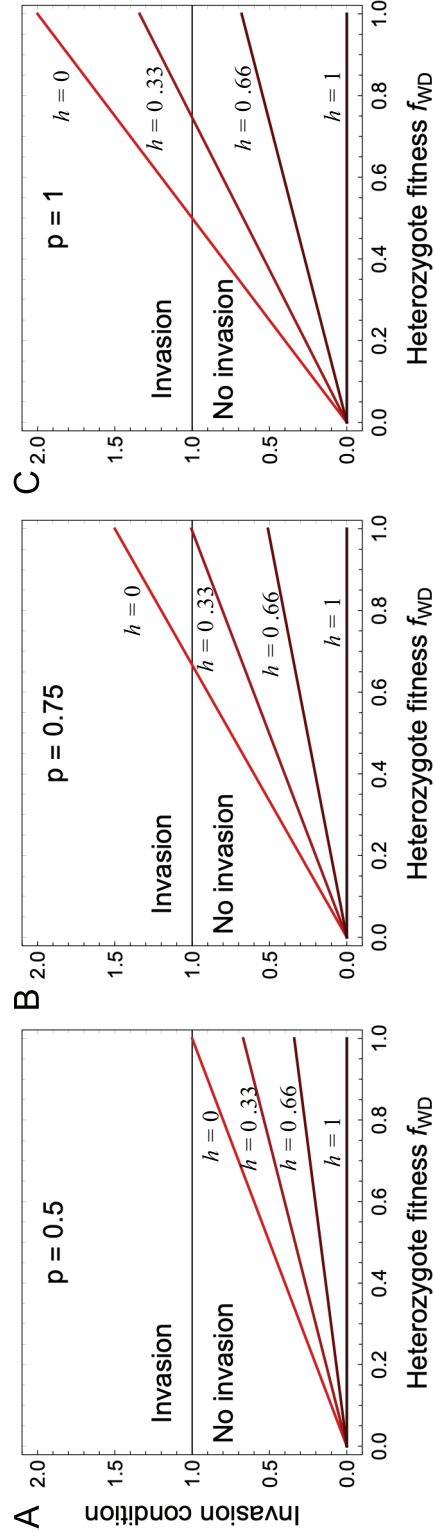


Figure A1: Invasion condition with varying mate choice bias (h) against heterozygote fitness f_{WD} . Here, invasion condition refers to the ability of the drive to invade a wild-type population from low frequency, with an invasion condition greater than 1 being necessary for success. A, Medea drive or no distortion, $p = 0.5$. The wild-type population cannot be invaded for any value of mate choice bias, h . B, Distortion-based gene drive with $p = 0.75$. The wild-type population can be invaded if there is no mate choice bias ($h = 0$) and $f_{WD} > 2/3$. C, Distortion-based gene drive with $p = 1$. The wild-type population can be invaded if mate choice bias is not very high (i.e., for $h = 0$ and $h = 0.33$).

APPENDIX B

Additional Figures

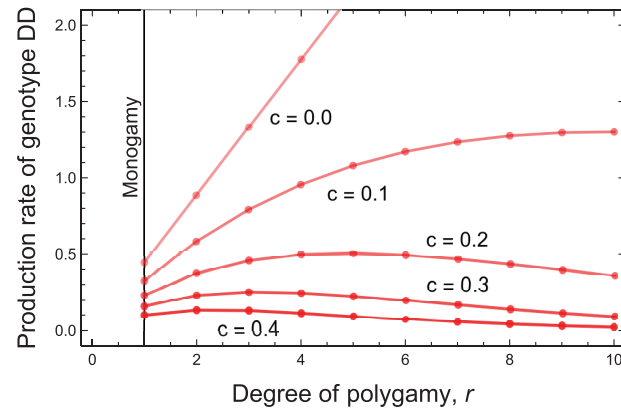


Figure B1: Effect on the rate of production of the DD genotype with increases in the degree of polygamy (r) for different fitness cost (c). We start from a population with an equal abundance of all three genotypes with 100% drive efficiency of the distortion-based gene drive for different fitness costs. In essence, we plotted equation (9) for varying r and c , keeping $x_{WW} = 1/3$, $x_{WD} = 1/3$, $x_{DD} = 1/3$, and $p = 1$. Increasing the fitness cost of the drive allele decreases the overall production of the DD genotype. For a moderate level of fitness cost, production of genotype DD first increases up to a point for species with a higher level of polygamy but then starts to decrease.

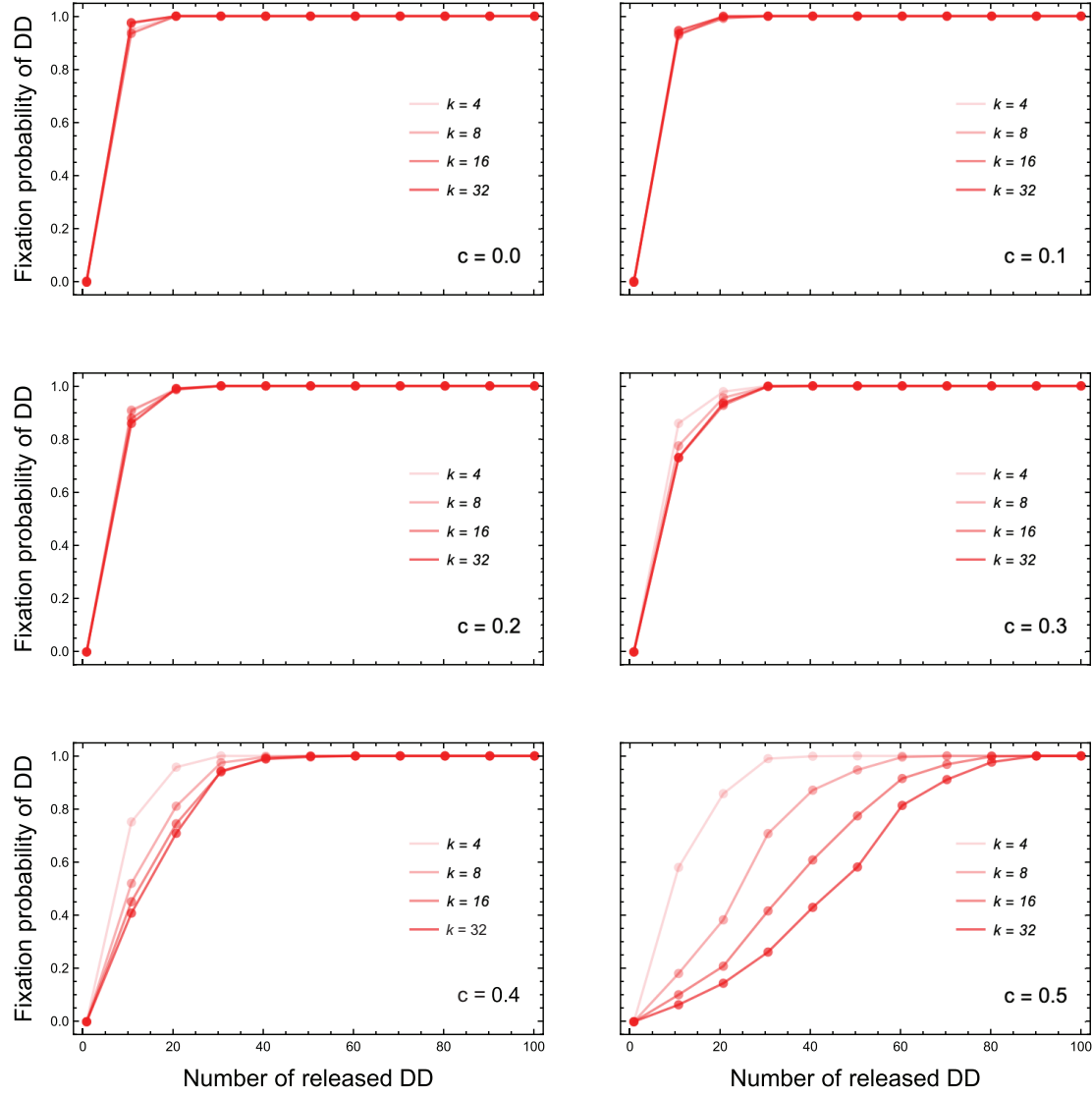


Figure B2: Effect of the network structure k on the fixation probability of DD for costless to costly drives (c). Plots show the fixation probability of drive homozygotes for networks with different average degree k for a variety of the number of released DD individuals. One DD individual is initially released in the population consisting only of WW. A generation consists of n death-birth steps. We see that for a costless drive, the degree of the network does not play a major role in the fixation probability, but as the cost c increases, the more connected networks need a larger release to reach the same probability of fixation as a weakly connected network. Hence, in a generation the whole population is updated on an average. All simulations were performed for a population size of $n = 100$ and 1,000 trials to estimate fixation probability. The distortion probability is set to $p = 0.95$.

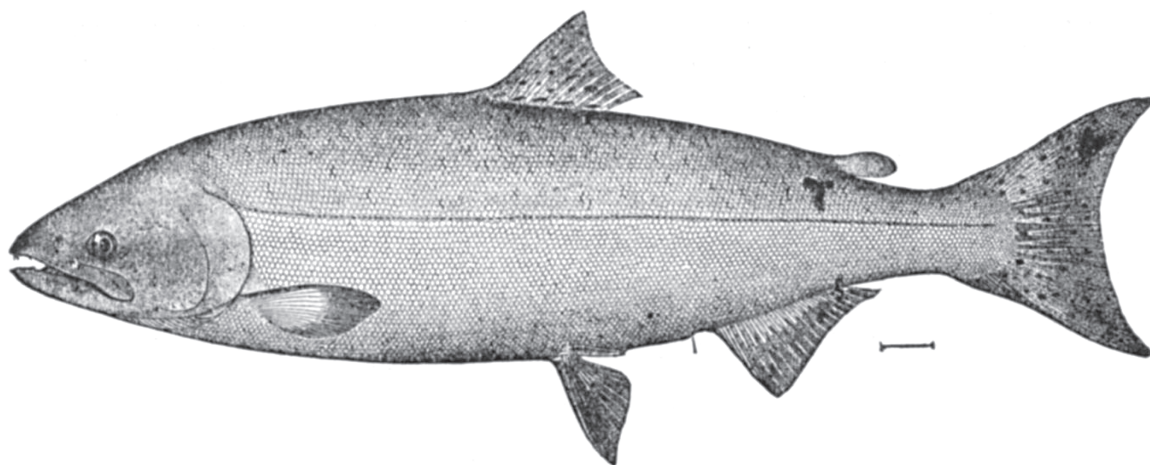
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Associate Editor: Tim Connallon
Editor: Erol Akçay



“The genus of Pacific salmons (*Oncorhynchus*) which is very closely related to *Salmo*, is represented by five species, all of which are more or less black-spotted, especially while sojourning in streams. They ascend the rivers falling into the North Pacific in Asia and North America.” Figured: “Quinnat Salmon (*Oncorhynchus chouicha*). Columbia River, Oregon.” From “Distribution and Some Characters of the Salmonidæ” by Tarleton H. Bean (*The American Naturalist*, 1888, 22:306–314).